

ENIGMA-PAK

in collaboration with the

National Centralized Repository for Alzheimer's Disease and Related Dementias



Biospecimen Collection, Processing, and Shipment Manual of Procedures

Version 04.2025



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1.0 Abbreviations

AD Alzheimer's Disease
CSF Cerebrospinal Fluid
DNA Deoxyribonucleic Acid

EDTA Ethylene Diamine Tetra-acetic Acid

GUID Globally Unique Identifier

IATA International Air Transport Association

NCRAD National Centralized Repository for Alzheimer's Disease and Related Dementias

PHI Protected Health Information

RBC Red Blood Cells

RCF Relative Centrifugal Force RPM Revolutions Per Minute UPS United Parcel Service

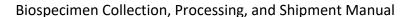
2.0 Purpose

The collection of biofluids is an important part of ENIGMA-PAK. The purpose of this manual is to provide study staff (PIs, study coordinators, phlebotomists) at the various study sites with instructions for collection and submission of biological samples for study visits. It includes instructions for biofluid submission to NCRAD located in Indianapolis at Indiana University.

Sites will collect and send the following samples to NCRAD:

- Plasma
- Buffy Coat (DNA Extraction)

This manual includes instructions for collection of blood, storage prior to shipping and shipping to NCRAD. These procedures are relevant to all study personnel responsible for processing specimens provided to NCRAD for the ENIGMA-PAK protocol.





3.0 NCRAD Information

3.1 NCRAD Contacts

Tatiana Foroud, PhD, Core Leader

Phone: 317-274-2218

Kelley Faber, MS, CCRC, Senior Project Manager

Phone: 317-274-7360 Email: kelfaber@iu.edu

Mica Gosnell, MS, Clinical Research Coordinator

Phone: 317-274-7423 Email: gosnellm@iu.edu

General NCRAD Contact Information

Phone: 1-800-526-2839 or 317-278-8413

Email: <u>alzstudy@iu.edu</u>
Website: <u>www.ncrad.org</u>

Sample Shipment Mailing Address

NCRAD

Indiana University School of Medicine 351 W. 10th St. TK-317

Indianapolis, IN 46202 Phone: 1-800-526-2839

3.2 NCRAD Hours of Operation

Indiana University business hours are from 8 AM to 5 PM Eastern Time, Monday through Friday.

Check the weather report to make sure impending weather events (blizzards, hurricanes, etc.) will not impact the shipping or delivery of the samples.



3.3 NCRAD Holiday Observations

| Date | Holiday |
|--------------------------------------|----------------------------|
| January 1 | New Year's Day |
| 3 rd Monday in January | Martin Luther King, Jr Day |
| 4 th Monday in May | Memorial Day |
| June 19 | Juneteenth |
| July 4 | Independence Day |
| 1 st Monday in September | Labor Day |
| 4 th Thursday in November | Thanksgiving |
| 4 th Friday in November | Friday after Thanksgiving |
| December 25 | Christmas Day |
| December 26-December 30 | Winter Break |

Please note that between December 24th and January 2nd, Indiana University will be open Monday through Friday for essential operations **ONLY** and will re-open for normal operations on January 2nd. If possible, biological specimens for submission to Indiana University should **NOT** be collected and shipped to Indiana University after the second week in December. Should it be necessary to ship blood samples for DNA extraction to Indiana University during this period, please contact the Indiana University staff before December 20th by e-mailing alzstudy@iu.edu, so that they can arrange to have staff available to process incoming samples. **Please see:** https://ncrad.org/contact/holiday-closures **for additional information.**

- Please note that courier services may observe a different set of holidays.
- Please be sure to verify shipping dates with your courier prior to any holiday.
- Weekend/holiday delivery must be arranged in advance with NCRAD staff.

4.0 Laboratory Collection

4.1 Site Required Equipment

The following materials and equipment are necessary for the processing of specimens at the collection site and are to be **supplied by the local site**:

- Personal Protective Equipment: lab coat, nitrile/latex gloves, safety glasses
- > Tourniquet
- Alcohol Prep Pad
- Gauze Pad
- Bandage
- > Butterfly needles and hub
- Microcentrifuge tube rack



- > Sharps bin and lid
- Wet Ice Bucket
- Wet ice
- Pelleted dry ice

In order to process samples consistently across all projects and ensure the highest quality samples possible, project sites must have access to the following equipment:

- Centrifuge capable of ≥ 2000 x g with refrigeration to 4°C
- > -80°C Freezer

In order to ship specimens, you must provide:

Pelleted dry ice

4.2 Biospecimens Sent to NCRAD

Guidelines for the processing, storage location, and timing of sample collection are listed in the following tables.

4.2.1 Biofluid Collection Schedule

| Sample Type | Visit |
|-------------------|-------|
| Plasma | Х |
| Buffy Coat | Х |

Consent forms must specify that any biological samples and de-identified clinical data may be shared with academic and/or industry collaborators through NCRAD. Recommended consent language can be found on the NCRAD website at: https://ncrad.org/bank-samples/sample-management/recommended-consent-language. A copy of the consent form for each participant should be kept on file by the site investigator.

4.2.2 Biofluid Collection Charts

| Collection Tube Drawn At | | Specimen Type | Aliquot Volume | Total Number of Aliquots | Shipping Temperature |
|---------------------------|------------|------------------|-----------------------------|--------------------------|-------------------------|
| 2 EDTA (Purple-Top) Blood | Each visit | Plasma | 1.5 ml plasma aliquots | Up to 7 | Frozen |
| Collection Tubes (10 ml) | Each visit | Buffy Coat | ~1.0 ml buffy coat aliquots | 1 | Frozen |



5.0 Specimen Collection Kits, Shipping Kits, and Supplies

ENIGMA-PAK will provide: 1) Blood sample collection kits for research specimens to be stored at NCRAD and the Frozen Shipment Kit. The kits include blood tubes, pipettes, labels, boxes for plasma, buffy coat aliquots, as well shipping materials to send biospecimens to NCRAD.





6.0 Blood Collection and Processing Procedures

- 6.1 Whole Blood Collection with 10 ml EDTA (Purple-Top) Tube for Plasma and Buffy Coat
 - 1. Store empty EDTA tubes at room temperature, $64^{\circ}F$ $77^{\circ}F$ (18 °C 25 °C) before use. Check expiration dates on all collection tubes before visit.
 - 2. Set centrifuge to 4°C to pre-chill before use.
 - 3. Using a blood collection set and a holder, collect blood into the EDTA (Purple-Top) Blood Collection Tube (10 ml) using your organizations's recommended procedure for standard venipuncture technique. The following techniques shall be used to prevent possible backflow:
 - a. Place participant's arm in a downward position.
 - b. Hold tube in a vertical position, below the participant's arm during blood collection.
 - c. Release tourniquet as soon as blood starts to flow into last collection tube.



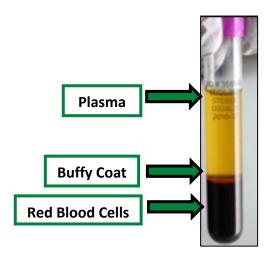
- d. Make sure tube additives do not touch stopper or end of the needle during venipuncture.
- 4. Allow at least 10 seconds for a complete blood draw to take place in each tube. Ensure that the blood has stopped flowing into the tube before removing the tube from the holder. The tube with its vacuum is designed to draw 10 ml of blood into the tube.
 - a. If complications arise during the blood draw, please note the difficulties. Do not attempt to draw an additional EDTA tube at this time. Process blood obtained in existing EDTA tube.
- 5. Immediately after blood collection, gently invert/mix (180 degree turns) the EDTA tube 8-10 times.
- 6. Immediately after inverting the EDTA tube, place it on wet ice until centrifugation begins.

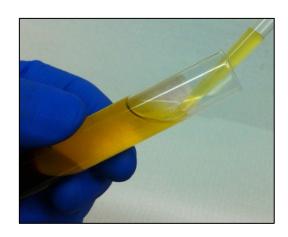


- 7. Centrifuge balanced tube for 10 minutes at 2000 x g at 4°C. It is critical that the tube be centrifuged at the appropriate speed and temperature to ensure proper plasma separation (see worksheet in Appendix A to calculate equivalent RPM for spin at 2000 x g).
 - a. While centrifuging, remember to record all times, temperatures and spin rates on the Biological Sample and Shipment Notification Form.
- 8. Record original volume drawn for each tube in spaces provided on the Biological Sample and Shipment Notification Form.
 - b. Plasma samples need to be spun, aliquoted, and placed in the freezer within 2 hours from the time of collection.
- 9. Record time aliquoted on the Biological Sample and Shipment Notification Form.



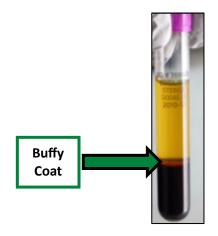
10. Remove the plasma by tilting the tube and placing the pipette tip along the lower side of the wall being careful not to agitate the packed red blood cells at the bottom of the collection tube.

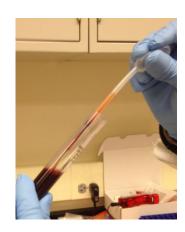




- 11. Each EDTA tube should yield, on average, 4-5 ml of plasma. Transfer plasma from both EDTA tubes into the 15 ml conical tube and gently invert 3 times. Aliquot 1.5 ml plasma per cryovial. Be sure to only place plasma in cryovials with purple caps and labeled with PLASMA labels. Place residual plasma (<1.5 ml) in the blue-capped cryovial. If a residual aliquot (<1.5 ml) is created, document the specimen number and volume on the Biological Sample and Shipment Notification Form.
- 12. Place the labeled cryovials in the 25 cell cryobox and place on pelleted dry ice. **Transfer to -80°C Freezer when possible**. Store all samples at **-80°C until shipped** to NCRAD on pelleted dry ice. Record time aliquots frozen and storage temperature of freezer on Biological Sample and Shipment Notification Form (Appendix B).
- 13. After plasma has been removed from the EDTA (Purple-Top) Blood Collection Tubes (10 ml), aliquot the buffy coat layer (in the top layer of cells, the buffy coat is mixed with RBCs-see figure) from one EDTA tube into a labeled, clear-capped cryovial using a micropipette. The buffy coat aliquot is expected to have a reddish color from the RBCs. Be sure to only place the buffy coat from one EDTA tube into each cryovial. Repeat this step for the second EDTA tube, placing this buffy coat into the second clear-capped cryovial.



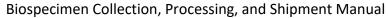






Buffy Coat Aliquot (clear-capped cryovial)

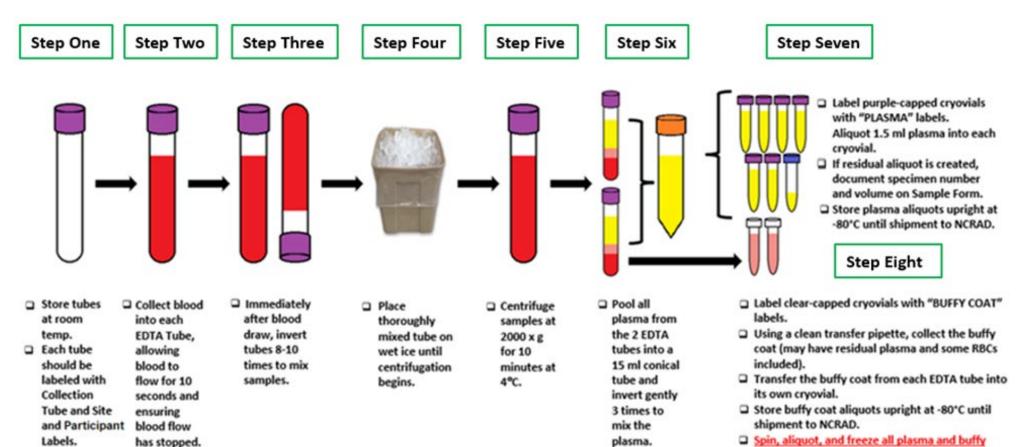
- 14. Dispose of collection tube with red blood cell pellet according to your site's guidelines for disposing of biomedical waste.
- 15. Record the specimen number and volumes of the EDTA tubes and corresponding buffy coat samples on the Shipping Manifest.
- 16. Place the labeled cryovials in the 25 cell cryobox and place on pelleted dry ice. **Transfer to -80°C Freezer when possible.** Store all samples at -80°C until shipped to NCRAD on pelleted dry ice. Record time aliquots frozen and storage temperature of freezer on Biological Sample and Shipment Notification Form.



coat aliquots within 2 hours of collection.



Plasma and Buffy Coat Preparation EDTA Purple-Top Tube (2 x 10 ml)





7.0 Packaging & Shipping Instructions

ALL study personnel responsible for shipping should be certified in biospecimen shipping. If you have difficulty finding biospecimen shipping training, please notify a NCRAD coordinator.

7.1 Frozen Packaging Instructions

The most important issue for shipping is to maintain the temperature of the samples. The frozen samples must never thaw; not even the outside of the tubes should be allowed to defrost. This is best accomplished by making sure the Styrofoam container is filled completely with pelleted dry ice.

Specimens being shipped to NCRAD should be considered as Category B UN3373 specimens and as such must be tripled packaged and compliant with IATA Packing Instructions 650. See the Latest Edition of the IATA Regulations for complete documentation.

Triple packaging consists of a primary receptacle(s), a secondary packaging, and a rigid outer packaging. The primary receptacles must be packed in secondary packaging in such a way that, under normal conditions of transport, they cannot break, be punctured, or leak their contents into the secondary packaging. Secondary packaging must be secured in outer packaging with suitable cushioning material. Any leakage of the contents must not compromise the integrity of the cushioning material or of the outer packaging.

7.1.1 NCRAD Packaging Instructions – Frozen Shipments

- 1. World Courier provides all shipment materials (boxes, dry ice, etc).
- 2. If possible, hold packaged samples in -80°C freezer until time of pick-up/drop-off. If storage in a -80°C freezer until pick-up is not possible, package samples no more than 4 hours before the expected pick-up time.
- 3. Notify NCRAD of shipment by emailing NCRAD coordinators at alzstudy@iu.edu. Attach the following to the email:
 - a. Completed Sample Forms (<u>Appendix B</u>) to the email notification (email NCRAD coordinator prior to shipment to receive sample form).
 - a. If email is unavailable, please call NCRAD and do not ship until you've contacted and notified NCRAD coordinators about the shipment in advance.
 - b. Tracking number from courier



4. Follow ENIGMA-PAK study and courier packaging instructions to ensure a sufficient amount of dry ice is packed in each shipper to ensure sample integrity upon arrival at NCRAD.

8.0 Appendices

Appendix A: Rate of Centrifuge Worksheet

Appendix B: NCRAD Blood Sample and Shipment Notification Form

Appendix C: ENIGMA PAK Shipping Manifest

Appendix D: Shipping Documentation

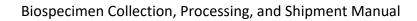


Biospecimen Collection, Processing, and Shipment Manual **Appendix A: Rate of Centrifuge Worksheet**

Please complete and return this form by email to the NCRAD Project Manager if you have any questions regarding sample processing. The correct RPM will be sent back to you.

| Submitter Information Name: Submitter e-mail: | Site: |
|--|--|
| Centrifuge Information Please answer the following questions about your ce | ntrifuge. |
| Centrifuge Type Fixed Angle Rotor: □ Swing Bucket Rotor: □ | |
| Radius of Rotation (mm): | |
| Determine the centrifuge's radius of rotation (in mm the centrifuge spindle to the bottom of the device who wing bucket rotor, measure to the middle of the bucket rotor. | hen inserted into the rotor (if measuring a |
| Calculating RPM from G-Force: $RCF = \left(\frac{RPM}{1,000}\right)^{2} \times r \times 1.118 \implies$ | $RPM = \sqrt{\frac{RCF}{r \times 1.118}} \times 1,000$ |
| RCF = Relative Centrifugal Force (G-Force) RPM = Rotational Speed (revolutions per minute) R= Centrifugal radius in mm = distance from the cent centrifuge | er of the turning axis to the bottom of |
| Comments: | |

Please send this form to NCRAD Study Coordinator at alzstudy@iu.edu





Appendix B: NCRAD Blood Sample and Shipment Notification Form

Please email the form on or prior to the date of shipment.

| To: Kelley Faber (NCRAD) Email: alzstudy@iu.edu Phone: 1-800-526-2839 | | | | | |
|---|----------------|----------------------------|-----------|--|--|
| From: | Trac | king #: | | | |
| Phone: | Phone: Email: | | | | |
| Study: ENIGMA-PAK | | | | | |
| Participant ID: | Sex | : M F Year of Birth: | | | |
| Blood Collection: | | | 1 | | |
| Date of Draw: | [MMDDYY] | Time of Draw: | [HHMM] | | |
| Date participant last ate: | [MMDDYY] | Time participant last ate: | [HHMM] | | |
| | | | _ | | |
| Blood Processing: | Plasma and Buf | fy Coat (EDTA Tube) | | | |
| EDTA #1 specimen number | | Original blood volume of | | | |
| (Last four digits): | | EDTA #1: | mL | | |
| EDTA #2 specimen number | | Original blood volume of | | | |
| (Last four digits): | N/A | EDTA #2: | mLN/A | | |
| Time spin started: | [HHMM] | Duration of centrifuge: | mins | | |
| Temp of centrifuge: | °C | Rate of centrifuge: | RCF (x g) | | |
| | | Number of 1.5 mL aliquots | _ | | |
| Time aliquoted: | (HHMM) | created (purple cap): | N/A | | |
| | | Specimen number of | | | |
| Volume of residual plasma aliquot | | residual plasma aliquot | | | |
| (less than 1.5 mL in blue cap): | mLN/A | (Last four digits): | N/A | | |
| Buffy coat #1 specimen number | | D (() 114 | 1 | | |
| (Last four digits): | | Buffy coat #1 volume: | mL | | |
| Buffy coat #2 specimen number (Last four digits): | N/A | Buffy coat #2 volume: | mL ∏N/A | | |
| , , , | | Storage temperature of | | | |
| Time aliquots frozen: | [HHMM] | freezer: | °C | | |
| Notes: | | | | | |



Appendix C: ENIGMA PAK Shipping Manifest

Please email the form on or prior to the date of shipment.

| | | | | | | | | Fall | | | | | | | | | |
|---------------|-------------|---------|----------------------|---------------|------------------------------------|----------|-------------|-------------------------|--|--|---------------------|-----------|---------------|------------------------|-----------|-----|----------|
| | | | | | | Plea | se complete | this form when sending: | SMA PAK Sample Sn Samples to NCRAD. T | ipments 'he contact information for ei form BEFORE the samples | mailing the form is | | | | | | |
| | | | | | | | in the | e box below. NCRAD wo | uld like to receive this | form BEFORE the samples | arrive. | | | | | | |
| | | | | | | | | | | | | | | | | | |
| | | | | | | | | | | | | , | | | | | |
| | | | | | | To: Kell | ey Faber | Fax: 317-278-1100 | Email: alzstudy@ | iu.edu | | | | | | | |
| | | | | | | Phone: | 1-800-526-2 | 839 | | | | | | | | | |
| | | | | | | | | | | | | J | | | | | |
| | | | | | | | | | | | | | | | | | |
| | | | | | | From:_ | | | Center/Site ID: | | | | | | | | |
| | | | | | | Phone: | | | Fax: | | | | | | | | |
| | | | | | | Email: | | | Date: | | | | | | | | |
| | | | | | | | | | | | | 1 | | | | | |
| | | | 0 | | D-1- D (1 | | | | | | | | | 01 | | | |
| | | | Sez (Male/Female/ | | Date Drawn (date of blood draw) | Time of | | Unit of | Time Spin | Duration of | Rate of | Time | Time Aliquots | Storage Temperature | | | |
| Individual ID | Sample Type | Barcode | Unknown) | Year of Birth | | Draw | Yolume | Measurement | Started | Centrifuge | Centrifuge | Aliquoted | Frozen | of Freezer | Tube Type | Boz | Position |
| | | | | | | | | | | | | | | | | | |
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Appendix D: Shipping Documents

CUSTOMS INVOICE

| Date: | 28th September 2024 |
|-----------|--|
| Shipper: | Dr. XXXXXXX, Assistant Professor, Department of Paediatrics & Child Health Attn: Ms. Aneeta Hotwani Manager Infectious Diseases Department of Pediatrics and Child Health The Aga Khan University Stadium Road Karachi74800, Pakistan Tel: 92 21 3486 xxx/92 21 34864990 E-mail: xxxxxxxx@aku.edu / aneeta.hotwani@aku.edu |
| Consignee | Dr.xxxxxxxxxxx c/o Dr. xxxxxx Address: Building : Rooom #: Street #: City : Country: Phone: Email: Alternative contact info: Phone #: Fax #: Mobile #: |
| • • • | The materials being shipped are Human Blood samples. Biological Substance, Category B UN 3373 Type of sample: Human whole Milk samples Quantity: xxx vials in xx Boxes Packed on dry Ice 01 Box This Shipment contains material obtained from humans that has not been inoculated with or exposed to any livestock or poultry disease agent. The material is not tissue culture Origin. The Materials is not zoonotic. The material is not from Non-human primates. For Research analysis only. The specimen is for Laboratory testing only. For Customs Purpose Only \$10 |

Mr. Abdul Khaliq Pesnani Consultant, Imports & Special Assignment CNIC# 42000-0500070-7 CELL# 0300-8224185 Khaliq.pesnani@aku.edu



Biospecimen Collection, Processing, and Shipment Manual **CDC Declaration**

| Shipper: | Dr. XXXXXXX, |
|----------|---|
| | Assistant Professor, Department of Paediatrics & Child Health |

Attn: Ms. Aneeta Hotwani Manager Infectious Diseases

Department of Pediatrics and Child Health

The Aga Khan University

Stadium Road Karachi74800, Pakistan *Tel: 92 21 3486 xxx/92 21 34864990*

E-mail: xxxxxxx@aku.edu / aneeta.hotwani@aku.edu

| Consignee | Dr.xxxxxxxxxxxxxx |
|-----------|-------------------|
| Consignee | DI.XXXXXXXXXX |

c/o Dr. xxxxxx

Address: Building: Rooom #:

Street #:____

City: Country:

Phone: Email:

Alternative contact info:

Phone #: _____ Fax #: _____

Mobile #:____

Contents: The materials being shipped are Human whole Milk samples.

Biological Substance, Category B UN 3373

Type of sample: Human whole Milk samples Quantity: 267 vials in 04 Boxes

Packed on dry Ice 01 Box

- This Shipment contains material obtained from humans that has not been inoculated with or exposed to any livestock or poultry disease agent.
- The material is not tissue culture Origin.
- The material is not zoonotic.
- The material is not from Non-human primates.

For Research analysis only. The specimen is for Laboratory testing only.

For Customs Purpose Only \$10

Mr. Abdul Khaliq Pesnani

Consultant, Imports & Special Assignment

CNIC# 42000-0500070-7

CELL# 0300-8224185

Khaliq.pesnani@aku.edu Date: 28th September 2024



Name of Cargo Agent:

Airway Bill No:

UNDERTAKING

This is to certify that shipment does not contain any unauthorized explosives, narcotics and firearms or hazardous material; I consent to a search of this shipment.

I am aware that this endorsement containing original signatures, along with other shipping documents will be retained on file until the shipment is delivered.

| Number of Cartons: Weight: | 1 Box Dry Ice Shipment | | | | | |
|-------------------------------|--|--|--|--|--|--|
| Commodities : | Human whole Milk samples Quantity: 267 vials in 04 Boxes | | | | | |
| | Packed on Dry Ice. | | | | | |
| | 01 Box Dry Ice Shipment | | | | | |
| Destination: | Dr.xxxxxxxxxxx | | | | | |
| | c/o Dr. xxxxxx | | | | | |
| | Address: Building : Rooom #: | | | | | |
| | Street #: | | | | | |
| | City: Country: | | | | | |
| | Phone: | | | | | |
| | Email: | | | | | |
| | Alternative contact info: | | | | | |
| | Phone #: Fax #: | | | | | |
| | Mobile #: | | | | | |

BNS Logistics (Pvt) Ltd.

Mr. Abdul Khaliq Pesnani Consultant, Imports & Special Assignment CNIC# 42000-0500070-7 CELL# 0300-8224185 Khaliq.pesnani@aku.edu



Biospecimen Collection, Processing, and Shipment Manual Dated: 28th September 2024

To Whom It May Concern:

| Dear Sir, |
|---|
| Reference to the above subject the undersigned of The Aga Khan University Hospital, Karachi, Department of Pediatrics and Child Health, an accredited for testing purpose, have to ship Human Blood samples Quantity: xxx vials in xx Boxes. Biological Substance, Category B UN 3373 Non-infectious, Non-hazardous packed with Dry Ice to xxxxxxxx For diagnostic and scientific analysis purpose. |
| The samples are meant solely for lab analysis purposes and are of no commercial value. |
| It is further certified that the samples are non-hazardous, non-infected and do not contain animal material cits by-products. |
| Thanking you for your cooperation and courtesy. |

With regards,

Mr. Abdul Khaliq Pesnani Consultant, Imports & Special Assignment CNIC# 42000-0500070-7 CELL# 0300-8224185 Khaliq.pesnani@aku.edu



Biospecimen Collection, Processing, and Shipment Manual Dated: 28th September 2024

Karachi AFU Airport Karachi Pakistan

| SUB: REQUEST FOR THE EXPORT THE SHIPMENT OF PATIENT SAMPLES MAWB# |
|---|
| Dear Sir, |
| Reference to the above subject the undersigned of The Aga Khan University Hospital, Karachi, Department of Pediatrics and Child Health, an accredited for testing purpose, have to ship Human Blood samples Quantity: xxx vials in xx Boxes. Biological Substance, Category B UN 3373 Non-infectious, Non-hazardous packed with Dry Ice to xxxxxxxx For diagnostic and scientific analysis purpose. |
| |
| This is to request you to kindly allow us to export these samples as they are temperature sensitive and are packed in dry ice and shall perish if delayed in shipping. |
| Thanking you for your cooperation and courtesy |
| With regards, |
| |

Mr. Abdul Khaliq Pesnani Consultant, Imports & Special Assignment CNIC# 42000-0500070-7 CELL# 0300-8224185 Khaliq.pesnani@aku.edu