

MCS-R01 Manual of Procedures Update: V04.14.2026

Section	Change
Throughout	Added "Important Note" about expired tubes to processing schematics.
Section 7.1.1	Removed email instructions for shipping information.
Section 8.0	Updated Electronic Sample and Shipping Forms Information.
Section 6.3 and Section 7.3	Added Electronic Data Capture Submission Instructions.
Section 3.0	Updated contact information.
Section 5.2	Changed "2 to 3 weeks" to "3 weeks."
Throughout	Updated Section Formatting and Numbering.
Section 6.1	Added specimen abbreviation table.
Section 6.1	Update labeling instruction and label diagrams.
Section 6.1	Added collection tube label and aliquot cryovial label language to label type summary table.
Section 6.1	Updated label format.
Section 3.3	Added winter break to Holiday Closures.
Section 7.1.1 Step 10	Specified to include Blood Sample and Shipment Notification Form on outside of Styrofoam container.
Section 7.1.1 Step 7	Specified maximum number of cryoboxes for shipping container.
Section 3.1	Added new URL for PACT study webpage on NCRAD website.
Section 6.1	Updated labels to reflect new format.
Throughout	Applied new template including change in colors, formats, and logo throughout.
Throughout	Alternate text applied to images and tables throughout.
Throughout	Replaced "dry ice" language with "pelleted dry ice" throughout.
Throughout	Replaced "subject" and "patient" language with "participant".
Section 3.1, Appendix B, and Section 7.1.1	Included alternate phone number in addition to NCRAD contact information.

Section	Change
Section 6.2 Step 4C	Updated Step C to clarify “Release tourniquet as soon as blood starts to flow into last collection tube”
Section 5.1	Removed “Biohazard Label” from individual supplies.
Section 6.2 Step 13	Included unused cryovials to disposal instruction.
Section 6.2 Step 8C	Language corrected from one-hour to two-hour window time



Biospecimen Collection, Processing, and Shipment Manual

Multicultural Community Dementia Screening

in collaboration with the

National Centralized Repository for Alzheimer's Disease and Related Dementias



**Biospecimen Collection, Processing, and Shipment Manual of
Procedures**

Version 04.14.2026

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1.0 Abbreviations

AD	Alzheimer's Disease
MCS	Multicultural Community Dementia Screening
DNA	Deoxyribonucleic Acid
EDTA	Ethylene Diamine Tetra-acetic Acid
GUID	Globally Unique Identifier
IATA	International Air Transport Association
NACC	National Alzheimer's Coordinating Center
NCRAD	National Centralized Repository for Alzheimer's Disease and Related Dementias
PHI	Protected Health Information
RBC	Red Blood Cells
RCF	Relative Centrifugal Force
RPM	Revolutions Per Minute

2.0 Purpose

The collection of biofluids is an important part of the Multicultural Community Dementia Screening (MCS) Study. The purpose of this manual is to provide study staff with instructions for collection and submission of biological samples for MCS study visits. It includes instructions for biofluid submission to NCRAD located in Indianapolis at Indiana University.

The following samples will be sent to NCRAD:

- Plasma
- Buffy Coat (DNA Extraction)

This manual includes instructions for collection of blood, fractionation of blood from collection tubes, aliquoting, labeling, storage prior to shipping, and shipping to NCRAD.

These procedures are relevant to all study personnel responsible for processing specimens provided to NCRAD for the MCS protocol.

3.0 NCRAD Information

3.1 NCRAD Contacts

Tatiana Foroud, PhD, Core Leader

Phone: 317-274-2218

Kelley Faber, MS, CCRC, Senior Project Manager

Phone: 317-274-7360

Email: kelfaber@iu.edu

Jordan Glogoza, BS, CCRC, Clinical Research Coordinator

Phone: 317-274-4307

Email: joglogoz@iu.edu

General NCRAD Contact Information

Phone: 1-800-526-2839/317-278-8413

Fax: 317-321-2003

Email: alzstudy@iu.edu

Website: www.ncrad.org

MCS Study Webpage: <https://ncrad.org/coordinate-studies/MCS>

Sample Shipment Mailing Address

MCS at NCRAD

Indiana University School of Medicine

351 W. 10th St. TK-342

Indianapolis, IN 46202

Phone: 1-800-526-2839/317-278-8413

3.2 NCRAD Hours of Operation

Indiana University business hours are from 8 AM to 5 PM Eastern Time, Monday through Friday.

Frozen samples must be shipped **Monday-Wednesday only**.

For packing and shipment details of samples, please refer to [Section 7.0](#) of this protocol.

Check the weather report to make sure impending weather events (blizzards, hurricanes, etc.) will not impact the shipping or delivery of the samples.

3.3 NCRAD Holiday Observations

Date	Holiday
January 1	New Year's Day
3 rd Monday in January	Martin Luther King, Jr Day
4 th Monday in May	Memorial Day
June 19	Juneteenth
July 4	Independence Day
1 st Monday in September	Labor Day
4 th Thursday in November	Thanksgiving
4 th Friday in November	Friday after Thanksgiving
December 25	Christmas Day
December 26-December 31	Winter Break

Please note that between December 24th and January 2nd, Indiana University will be open Monday through Friday for essential operations **ONLY** and will re-open for normal operations on January 2nd. If possible, biological specimens for submission to Indiana University should **NOT** be collected and shipped to Indiana University after the second week in December. Should it be necessary to ship blood samples for DNA extraction to Indiana University during this period, please contact the Indiana University staff before December 20th by e-mailing alzstudy@iu.edu, so that they can arrange to have staff available to process incoming samples. Please see: <https://ncrad.org/contact/holiday-closures> for **additional information**.

- Please note that courier services may observe a different set of holidays.
- Please be sure to verify shipping dates with your courier prior to any holiday.
- **Weekend/holiday delivery must be arranged in advance with NCRAD staff.**

4.0 MCS Laboratory Collection

4.1 Site Required Equipment

The following materials and equipment are necessary for the processing of specimens at the collection site and are to be **supplied by the local site**:

- Personal Protective Equipment: lab coat, nitrile/latex gloves, safety glasses
- Tourniquet
- Alcohol Prep Pad
- Gauze Pad
- Bandage
- Butterfly needles and hub
- Microcentrifuge tube rack
- Sharps bin and lid
- Wet Ice Bucket
- Wet ice
- Pelleted dry ice

In order to process samples consistently across all projects and ensure the highest quality samples possible, project site must have access to the following equipment:

- Centrifuge capable of $\geq 2000 \times g$ with refrigeration to 4°C
- -80°C Freezer

In order to ship specimens, you must provide:

- Pelleted dry ice (approximately 45 lbs per shipment)

4.2 Biospecimens Sent to NCRAD

Samples are to be submitted according to the shipping methods outlined in [Section 7.0](#). Guidelines for the processing, storage location, and timing of sample collection are listed in the tables below.

4.2.1 Biofluid Collection Schedule

Biospecimen Collection Table

Biospecimen	All Visits
Plasma	X
Buffy Coat (DNA)	X

Whole blood is collected in two collection tubes (two 10 ml purple-top EDTA tubes) for shipment to NCRAD. The 10 ml EDTA tubes are processed locally into

Biospecimen Collection, Processing, and Shipment Manual
 plasma and buffy coat fractions; they are then aliquoted, frozen at the study site, and shipped to NCRAD.

Consent forms must specify that any biological samples and de-identified clinical data may be shared with academic and/or industry collaborators through NCRAD. Recommended consent language can be found on the NCRAD website at: https://ncrad.org/recommended_consent_language.html. A copy of the consent form for each participant should be kept on file by the site investigator.

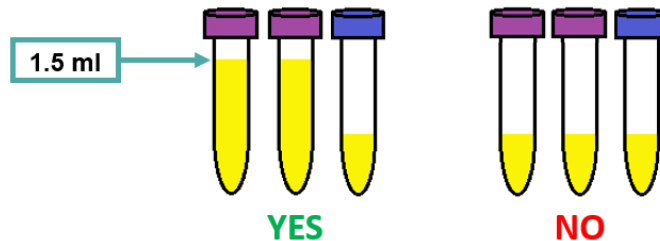
4.2.2 Biofluid Collection Charts

Collection Tube	Drawn At	Specimen Type	Aliquot Volume	Total Number of Aliquots	Shipping Temperature
2 EDTA (Purple-Top) Blood Collection Tubes (10 ml)	All Visits	Plasma	1.5 ml plasma aliquots	Up to 7	Frozen
	All Visits	Buffy Coat	~1.0 ml buffy coat aliquots	2	Frozen

4.3 Filling Cryovials

In order to ensure that NCRAD receives a sufficient amount of sample for processing and storage, and to avoid cracking of the tubes prior to shipment, each aliquot tube should be filled to the assigned volume after processing is completed (refer to detailed processing instructions for average yield per sample). Over-filled tubes may burst once placed in the freezer, resulting in a loss of sample.

Aliquot the remaining biologic material as the residual volume and ship to NCRAD. Ship *all* material to NCRAD. Fill as many aliquot tubes as possible. For example, if 2.7 ml of a plasma sample is obtained, fill 1 cryovial with 1.5 ml, and one additional cryovial with the remaining 1.2 ml.



Please note: It is critical for the integrity of future studies using these samples that study staff note if an aliquot tube contains a residual volume (anything under 1.5 ml). Please highlight that the aliquot contains a small volume by utilizing the blue cryovial cap provided in each kit. Please record the last four digits of the residual aliquot on the Biological Sample and Notification Form.

If there are any unused cryovials, **please do not send the empty cryovials to NCRAD. Our lab charges the study for the destruction of these empty tubes.**

These unused cryovials (ensure labels are removed) can be saved as part of a supplemental supply at your site or the cryovials can be disposed of per your site’s requirements.

To assist in the preparation and aliquoting of samples, colored caps are used for the aliquot tubes. The chart below summarizes the association between cap color and type of aliquot.

Cap Color	Sample Type
Lavender	Plasma
Clear	Buffy Coat
Blue	Residual sample plasma

5.0 Specimen Collection Kits, Shipping Kits, and Supplies

NCRAD will provide: 1) Blood sample collection kits for research specimens to be stored at NCRAD, the Blood Supplemental Supply Kit, the Frozen Shipment Kit and 2) clinical lab supplies (with the exception of pelleted dry ice and equipment supplies listed in [Section 5.1](#)). The provided materials include blood tubes, pipettes, boxes for plasma/buffy coat aliquots, as well as partially completed shipping labels to send materials to NCRAD. Kit number labels, site and PTID labels, collection tube labels, and aliquot labels will all be provided by NCRAD. Details regarding the blood kits are found in this Manual of Procedures. Collection tube and aliquot labels will be preprinted with study information specific to the type of sample being drawn. Ensure that all tubes are properly labeled during processing and at the time of shipment according to [Section 6.1](#).

5.1 NCRAD Specimen Collection Kit Contents

Collection kits contain the following (for each participant) and provide the necessary supplies to collect samples from a given participant. Do not replace or supplement any of the tubes or kit components provided with your own supplies unless you have received approval from the NCRAD Study team to do so. Please store all kits at room temperature until use.

MCS Blood Kit

Quantity	MCS Kit Components
2	EDTA (purple-top) blood collection tube (10 ml)
1	Centrifuge Tube, 15 ml
6	Cryovial (2.0 ml) with purple cap
1	Cryovial (2.0 ml) with blue cap
2	Cryovial (2.0 ml) with clear cap
9	Preprinted Cryovial Labels
5	Preprinted Kit Number Label
3	Label for handwritten Participant ID
1	Cryovial box (holds up to 25 cryovials)
3	Disposable graduated transfer pipettes (3 ml)
1	Resealable Bag
1	Resealable small poly bag (4" x 6")

MCS Supplemental Supply Kit

Quantity	MCS Supplemental Supply Kit Components
4	EDTA (purple-top) blood collection tube (10 ml)
12	Cryovial (2.0 ml) with purple cap
2	Cryovial (2.0 ml) with blue cap
4	Cryovial (2.0 ml) with clear cap
2	Conical tubes (50.0 ml)
5	Label for handwritten Participant ID
2	Cryovial box (holds up to 25 cryovials)
6	Disposable graduated transfer pipettes (3 ml)
1	Resealable bag
6	Resealable small poly bag (4" x 6")

NCRAD Frozen Shipping Supply Kit

Quantity	Frozen Shipping Kit Components for Blood-Based Biomarkers
8	Plastic Biohazard bag with absorbent sheet (small)
1	UPS return airbill and pouch
1	Shipping box/Styrofoam container
1	Warning label packet (UN3373, Fragile, Dry Ice)
1	Resealable bag

Quantities	Items Available upon request within the NCRAD kit module
By Request	Cryovial box (holds up to 25 cryovials)
By Request	Cryovial (2.0 ml) with purple cap
By Request	Cryovial (2.0 ml) with blue cap
By Request	Cryovial (2.0 ml) with clear cap
By Request	UPS return airbill
By Request	UPS Laboratory Pak
By Request	Shipping container for dry ice shipment (shipping and Styrofoam box)
By Request	Styrofoam shipping containers (11"x 9"x 8", 1 1/2" wall)
By Request	Plastic biohazard bag with absorbent sheet (small)
By Request	Disposable graduated transfer pipette (3 ml)
By Request	EDTA (Purple-Top) Blood Collection Tube (10 ml)
By Request	Warning label packet (UN3373, Fragile, Biohazard, Dry Ice)
By Request	UN3373 label
By Request	Dry ice shipping label
By Request	Fine Point Permanent Markers
By Request	Participant ID Labels

5.2 Kit Supply to Study Site

Each site will be responsible for ordering and maintaining a steady supply of kits from NCRAD. We advise sites to keep a supply of each kit type available. Be sure to check your supplies and order additional materials before you run out or supplies expire so you are prepared for study visits. Please go to: <https://redcap.link/MCSKitRequest> to request additional kits and follow the prompts to request the desired supplies.

Please allow **THREE weeks** for kit orders to be processed and delivered.

6.0 Blood Collection and Processing Procedures

6.1 Labeling Samples

*****Important Note*****

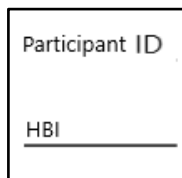
In order to ensure the highest quality samples are collected, it is essential to follow the specific collection and shipment procedures detailed in the following pages. Please read the following instructions first before collecting any specimens. Have all your supplies and equipment out and

Label Type Summary

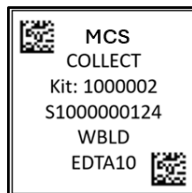
1. Kit Number Label
2. Participant ID Label
3. Collection Tube Label
4. Aliquot Cryovial Label



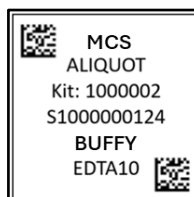
1. **Kit Number Labels** tie together all specimens collected from one participant at one visit. Place one kit number label on each cryobox.



2. **Participant ID Labels** are used to document the individual's unique study participant ID and the blood processing staff initials. Place one label on each blood collection tube.



3. **Collection Tube Labels** include the collection specimen type, kit number, and specimen barcodes. Place one collection tube label on each 10 ml EDTA blood collection tube.



4. **Aliquot Cryovial Labels** includes the processed specimen type, on each aliquot cryovial with corresponding specimen type. Place one aliquot cryovial label on corresponding specimen 2 ml cryovial.

Important Note

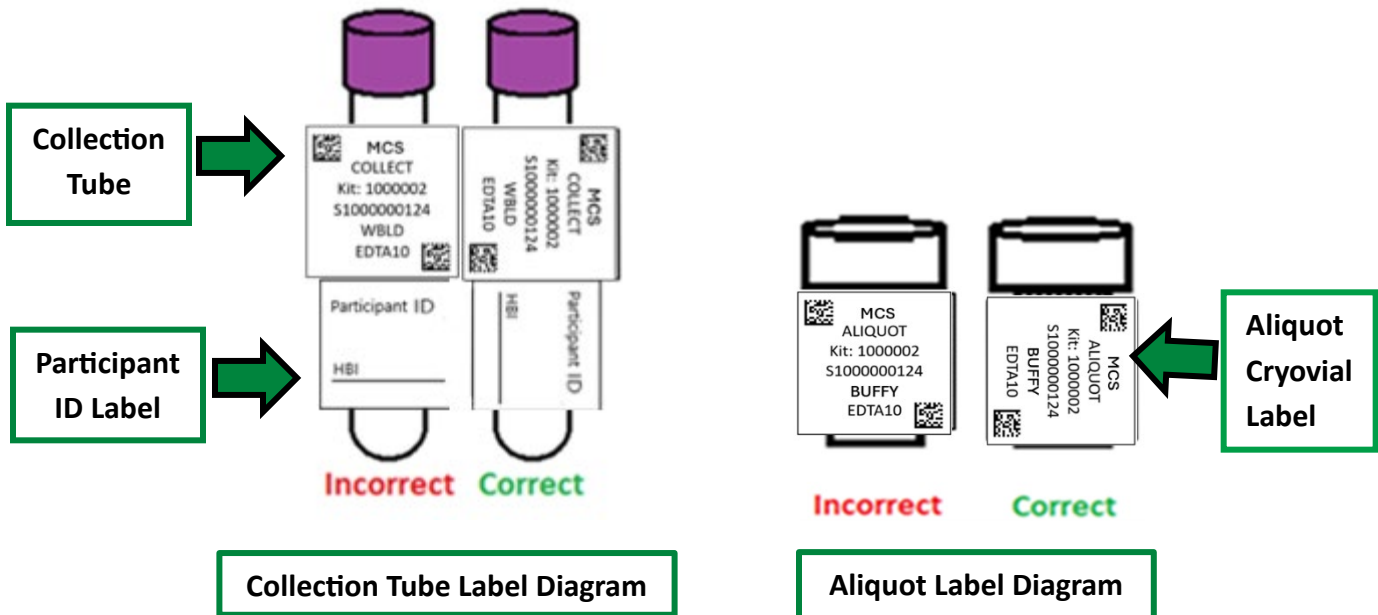
Each collection tube will contain two labels: the collection tube label and the Participant ID Label. Be sure to place labels in the same configuration consistently among tubes, with the barcoded label near the top of the tube and the handwritten Participant ID label.

Each aliquot cryovial will contain one label: the aliquot label is specimen specific and contains a unique barcode and number. Be sure that the specimen type, aliquot cap color, and specimen label correspond according to protocol.

In order to ensure the label adheres properly and remains on the tube, please follow these instructions:

- Place cryovial labels on **ALL** cryovials **BEFORE** sample collection. This should help to ensure the label properly adheres to the tube before exposure to moisture or different temperatures.
- Using a fine point permanent marker, fill-in and place the participant ID labels on the EDTA (purple-top) tubes **BEFORE** sample collection. These labels are placed on collection tubes in addition to the collection tube label.
- The collection tube labels contain a 2D barcode on the left-hand side of the label. Place this barcode toward the tube cap.
- Place label **horizontally** on the tube (wrapped around sideways if the tube is upright).

Take a moment to ensure the label is **completely adhered** to each tube. It may be helpful to roll the tube between your fingers after applying the label.



6.2 Whole Blood Collection with 10 ml EDTA (Purple-Top) Tube for Plasma and Buffy Coat

1. Store empty EDTA tubes at room temperature, 64°F - 77°F (18 °C – 25 °C) before use. Verify all supplies have not expired before use.
2. Set centrifuge to 4°C to pre-chill before use.
3. Place completed Participant ID Label and preprinted **PLASMA** Collection Tube Label on the purple-top EDTA tube. Place preprinted **PLASMA** Aliquot Labels on the six 2 ml cryovial tubes with purple caps and one 2 ml cryovial tube with blue cap (if necessary, for residual). Place preprinted **BUFFY COAT** cryovial label on the 2 ml cryovials with Clear caps.
4. Using a blood collection set and a holder, collect blood into the **EDTA (Purple-Top) Blood Collection Tube (10 ml)** using your institution's recommended procedure for standard venipuncture technique.

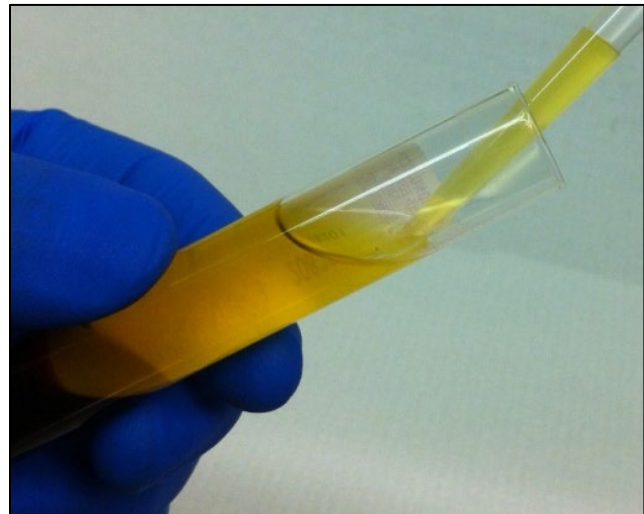
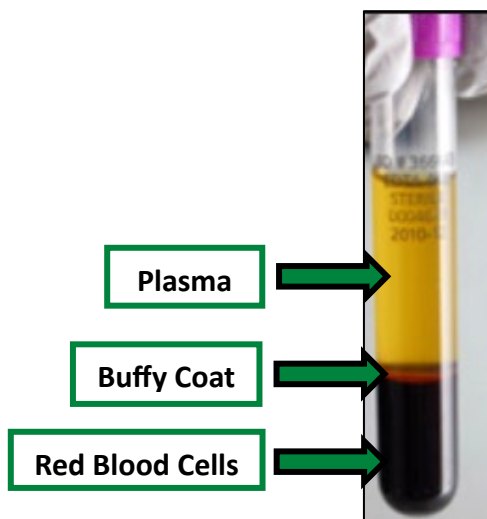
The following techniques shall be used to prevent possible backflow:

- a. Place participant's arm in a downward position.
 - b. Hold tube in a vertical position, below the participant's arm during blood collection.
 - c. Release tourniquet as soon as blood starts to flow into last collection tube.
 - d. Make sure tube additives do not touch stopper or end of the needle during venipuncture.
5. Allow at least 10 seconds for a complete blood draw to take place in each tube. **Ensure that the blood has stopped flowing into the tube before removing the tube from the holder.** The tube with its vacuum is designed to draw 10 ml of blood into the tube.
 - a. If complications arise during the blood draw, please note the difficulties on the 'Biological Sample and Shipment Notification Form'. Do not attempt to draw an additional EDTA tube at this time. Process blood obtained in existing EDTA tube.
 6. Immediately after blood collection, gently invert/mix (180 degree turns) the EDTA tube 8-10 times.
 7. Immediately after inverting the EDTA tube, place it on wet ice until centrifugation begins.

8. Centrifuge balanced tubes for 10 minutes at 2000 x g at 4°C. **It is critical that the tubes be centrifuged at the appropriate speed and temperature to ensure proper plasma separation (see worksheet in [Appendix A](#) to calculate equivalent RPM for spin at 2000 x g).**
 - a. While centrifuging, remember to record all times, temperatures and spin rates on the Biological Sample and Shipment Notification Form.
 - b. Record original volume drawn for each tube in spaces provided on the Biological Sample Shipment and Notification Form.
 - c. Plasma samples need to be spun, aliquoted, and placed in the freezer within 2 hour from the time of collection.
 - d. Record time aliquoted on the Biological Sample Shipment and Notification Form.

9. Remove the plasma by tilting the tube and placing the pipette tip along the lower side of the wall being careful not to agitate the packed red blood cells at the bottom of the collection tube.

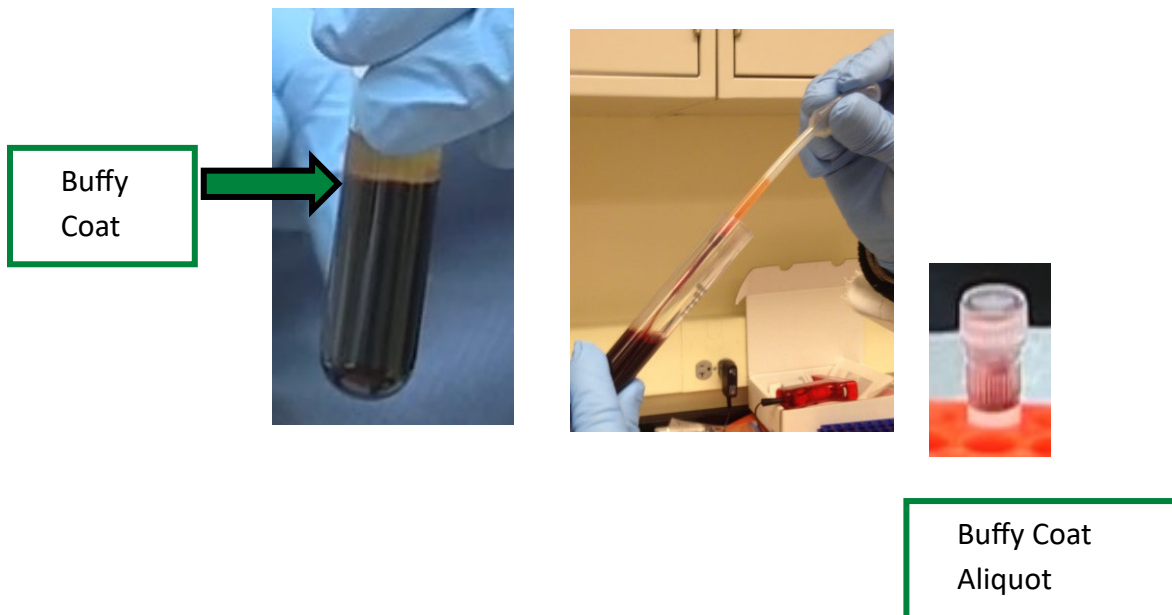
10. Each EDTA tube should yield, on average, 4-5 ml of plasma. Transfer plasma from both EDTA tubes into the 15 ml conical tube and gently invert 3 times. Aliquot 1.5 ml plasma per cryovial. Be sure to only place **plasma** in cryovials with Purple caps and labeled with **PLASMA** labels. Place residual plasma (<1.5 ml) in the blue-capped cryovial. **If a residual aliquot (<1.5 ml) is created, document the specimen number and volume on the Biological Sample and Shipment Notification Form.**



NOTE: When pipetting plasma from the EDTA tube into the 15 ml conical tube, be very careful to pipette the plasma top layer only, leaving the buffy coat and the red blood cell layers untouched.

11. Place the labeled cryovials in the 25 cell cryobox and place on pelleted dry ice. **Transfer to -80°C Freezer when possible.** Store all samples at **-80°C until shipped** to NCRAD on pelleted dry ice. Record time aliquots placed in freezer and storage temperature of freezer on Biological Sample Shipment and Notification Form.

12. After plasma has been removed from the EDTA (Purple-Top) Blood Collection Tubes (10 ml), aliquot the buffy coat layer from one EDTA tube into a labeled, Clear-capped cryovial using a pipette. The buffy coat aliquot is expected to have a reddish color from the RBCs. Be sure to only place the buffy coat from one EDTA tube into each cryovial. Repeat this step for the second EDTA tube, placing this buffy coat into the second Clear-capped cryovial.



13. Dispose of collection tube with red blood cell pellet and unused cryovials according to your site’s guidelines for disposing of biomedical waste.

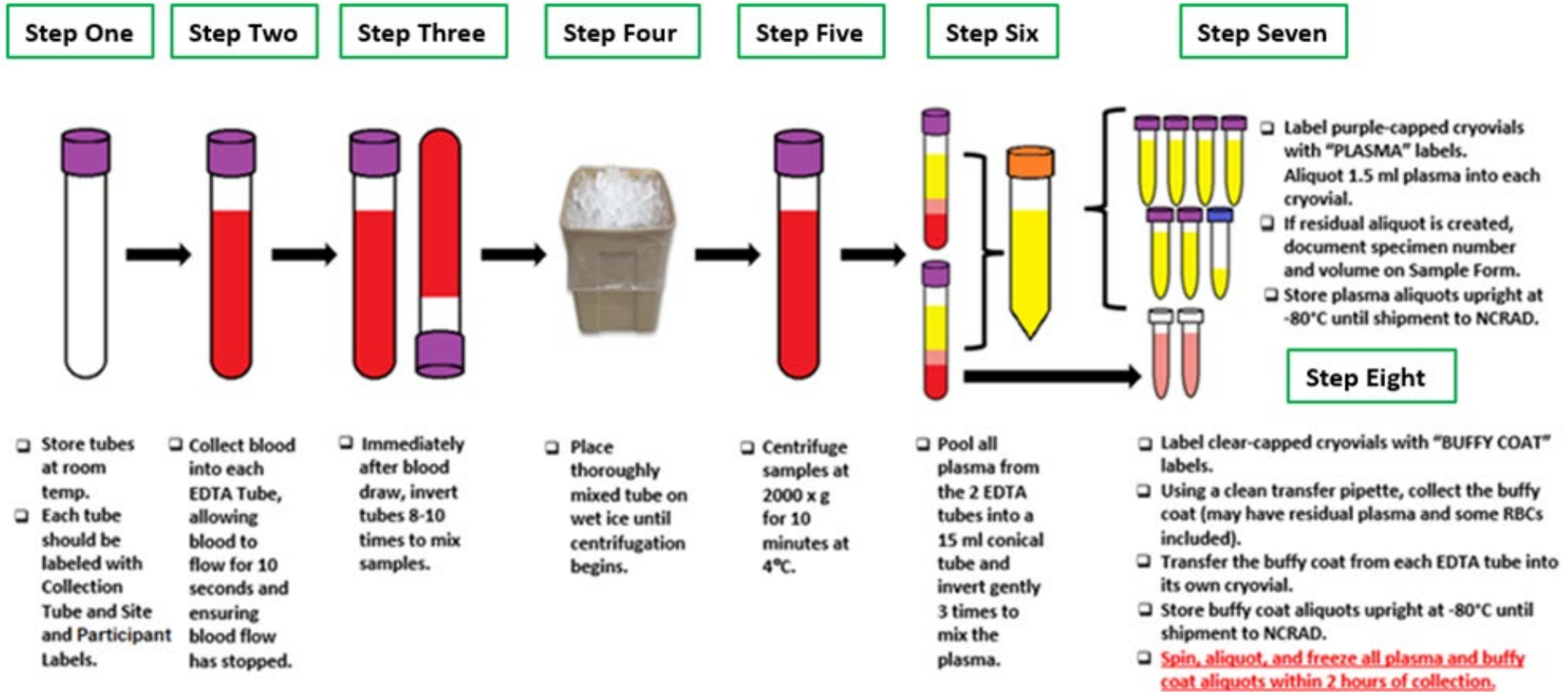
14. Record the specimen number and volumes of the EDTA tubes and corresponding buffy coat samples on the Biological Sample Shipment and Notification Form.

15. Place the labeled cryovials in the 25 cell cryobox and place on pelleted dry ice. **Transfer to -80°C Freezer when possible.** Store all samples at -80°C until shipped to NCRAD on pelleted dry ice. Record time aliquots placed in freezer and storage temperature of freezer on Biological Sample and Shipment Notification Form.



Plasma Aliquots (up to 7 possible) and Buffy Coats (2)

Plasma and Buffy Coat Preparation EDTA Purple-Top Tube (2 x 10 ml)



Important Note: Ensure all tubes are NOT expired prior to collection and processing of samples.

6.3 Sample Processing Electronic Data Capture Submission Instructions

1. After sample collection is completed, click here: <https://redcap.uits.iu.edu/MCSSampleForm> to report the data electronically. This data is required to be reported promptly after specimens have been processed.
2. Enter all required information, including:
 - a. Site information
 - b. Participant information
 - c. Visit information
 - d. Collection volumes
 - e. Processing details
 - f. Aliquoting information
 - g. Freezing time
3. If applicable, document any deviations from the protocol that occurred during collection, processing, or freezing in the “Notes” section.
4. Click “Continue to Review.”
5. You will be directed to a Condensed Form. Review the form carefully to verify that all information is correct
6. Click “Submit.”
You will automatically receive an email containing copies of both the Sample Form and the Condensed Form. Save the Condensed Form as a *PDF* and *print* a copy for your records.

7.0 Packaging & Shipping Instructions

ALL study personnel responsible for shipping should be certified in biospecimen shipping. If you have difficulty finding biospecimen shipping training, please notify a NCRAD coordinator.

In addition to tracking and reconciliation of samples, the condition and amount of samples received are tracked by NCRAD for each sample type. Investigators and clinical coordinators for each project are responsible to ensure the requested amounts of each fluid are collected to the best of their ability and that frozen samples are packed with sufficient amounts of pelleted dry ice to avoid thawing in the shipment process.

7.1 Frozen Packaging Instructions

The most important issue for shipping is to maintain the temperature of the samples. The frozen samples must never thaw; not even the outside of the tubes

Biospecimen Collection, Processing, and Shipment Manual should be allowed to defrost. This is best accomplished by making sure the Styrofoam container is filled completely with pelleted dry ice.

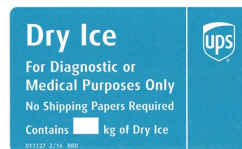
*****Important Note*****
FROZEN SAMPLES MUST BE SHIPPED MONDAY-WEDNESDAY ONLY!

Specimens being shipped to NCRAD should be considered as Category B UN3373 specimens and as such must be tripled packaged and compliant with IATA Packing Instructions 650. *See the Latest Edition of the IATA Regulations for complete documentation.*

Triple packaging consists of a primary receptacle(s), a secondary packaging, and a rigid outer packaging. The primary receptacles must be packed in secondary packaging in such a way that, under normal conditions of transport, they cannot break, be punctured, or leak their contents into the secondary packaging. Secondary packaging must be secured in outer packaging with suitable cushioning material. Any leakage of the contents must not compromise the integrity of the cushioning material or of the outer packaging.

***** Packing and Labeling Guidelines *****

- The primary receptacle (cryovial) must be leak proof and must not contain more than 1L total.
- The secondary packaging (biohazard bag) must be leak proof and if multiple blood tubes are placed in a single secondary packaging, they must be either individually wrapped or separated to prevent direct contact with adjacent blood tubes.
- Absorbent material must be placed between the primary receptacle and the secondary packaging. The absorbent material should be of sufficient quantity in order to absorb the entire contents of the specimens being shipped. Examples of absorbent material are paper towels, absorbent pads, cotton balls, or cellulose wadding.
- A shipping manifest of specimens being shipped must be included between the secondary and outer packaging.
- The outer shipping container must display the following labels:
 - ✓ Sender’s name and address
 - ✓ Recipient’s name and address
 - ✓ Responsible Person
 - ✓ The words “Biological Substance, Category B”
 - ✓ UN3373
 - ✓ UPS Dry Ice label and net weight of dry ice contained



7.1.1 NCRAD Packaging Instructions – Frozen Shipments

1. If possible, hold packaged samples in -80°C freezer until time of UPS pick-up/drop-off. If storage in a -80°C freezer until UPS pick-up is not possible, package samples no more than 4 hours before the expected pick-up time.
2. Contact UPS to confirm service is available and schedule package to be picked up.
3. Place the cryovial boxes containing frozen samples into a biohazard bag.
4. As the cryovial box is placed in the plastic biohazard bag, do NOT remove the absorbent material found in the bag. Seal according to the instructions on the bag.



5. Place approximately 2-3 inches of pelleted dry ice in the bottom of the Styrofoam shipping container.
6. Place the biohazard bags into the provided Styrofoam-lined shipping container on top of the pelleted dry ice. The large shipping container holds up to 8 cryoboxes. Please ensure that cryovial boxes are placed so the cryovials are upright in the shipping container.

7. Fully cover the biohazard bags containing the cryovial boxes tubes with approximately 2 inches of pelleted dry ice.
8. After the samples have been placed into the shipping container, completely fill the inner Styrofoam with pelleted dry ice pellets to ensure the frozen state of the specimens during transit.
9. Replace the lid on the Styrofoam carton. Place the completed Condensed Sample and Shipment Notification Form in the package on top of the outside of Styrofoam lid for each participant specimen, and close and seal the outer cardboard shipping carton with packing tape.
10. Complete the UPS Dry Ice Label with the following information:
 - a. Net weight of pelleted dry ice in kg (must match amount on the airbill)
 - b. Do not cover any part of this label with other stickers, including preprinted address labels.
11. Apply all provided warning labels and the preprinted UPS return airbill to the outside of package, taking care not to overlap labels.

*****Important Note*****

Complete the required fields on the UPS Dry Ice label or UPS may reject or return your

12. If possible, hold packaged samples in -80°C freezer until time of UPS pick-up/drop-off. If storage in a -80°C freezer until UPS pick-up is not possible, package samples no more than 4 hours before the expected pick-up time.
13. Specimens should be sent to the below address via UPS Next Day Air. Frozen shipments should be sent **Monday through Wednesday ONLY** to avoid shipping delays on Thursday or Friday.

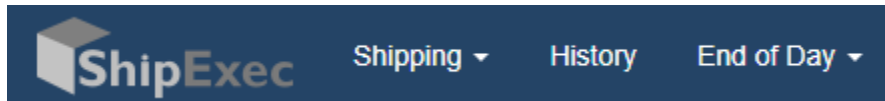
MCS at NCRAD
Indiana University School of Medicine
351 W. 10th St. TK-217
Indianapolis, IN 46202

14. Use UPS tracking to ensure the delivery occurs as scheduled and is received by NCRAD.

7.2 Frozen Shipping Instructions

1. Log into the ShipExec Thin Client at kits.iu.edu/UPS.
 - a. If a new user or contact needs access, please reach out to your study contact for access.

2. Click “Shipping” at the top of the page and select “Shipping and Rating”.



3. Select your study from the “Study Group” drop down on the right side of the main screen. Choosing your study will automatically filter the address book to only addresses within this study.
4. Click on the magnifying glass icon in the “Ship From” section to search for your shipping address.

Ship From

Company	<input style="width: 80%; height: 20px; border: 1px solid #ccc;" type="text"/>
Contact	<input style="width: 80%; height: 20px; border: 1px solid #ccc;" type="text"/>
Address 1	<input style="width: 80%; height: 20px; border: 1px solid #ccc;" type="text"/>
Address 2	<input style="width: 80%; height: 20px; border: 1px solid #ccc;" type="text"/>
Address 3	<input style="width: 80%; height: 20px; border: 1px solid #ccc;" type="text"/>
City	<input style="width: 80%; height: 20px; border: 1px solid #ccc;" type="text"/>
State/Province	<input style="width: 80%; height: 20px; border: 1px solid #ccc;" type="text"/>
Postal Code	<input style="width: 80%; height: 20px; border: 1px solid #ccc;" type="text"/>
Country/Territory	<input style="width: 80%; height: 20px; border: 1px solid #ccc;" type="text"/>
Phone	<input style="width: 80%; height: 20px; border: 1px solid #ccc;" type="text"/>

- a. Search by Company (site), Contact (name), or Address 1 (first line of your site’s street address). Click Search.
- b. Click Select to the left of the correct contact information.

5. Verify that both the shipping information AND study reference are correct for this shipment.
 - a. If wrong study contact or study reference, click Reset in the bottom right of the screen to research for the correct information.

6. Enter Package Information
 - a. Frozen shipments
 - i. Enter the total weight of your package in the “Weight” field.
 - ii. Enter the dry ice weight in the “Dry Ice Weight” field.
 - iii. If the “Dry Ice Weight” field is higher than the “Weight” field, you will receive an error message after clicking Ship and need to reenter these values.
 - b. Click Ship in the bottom right of the page when complete.

7. If your site does not already have a daily UPS pickup, you can schedule one here.
 - a. Click the blue Pickup Request button. Enter the earliest pickup time and latest pickup time in 24-hr format.
 - b. Give a name & phone number of someone who the UPS driver can call if having issues finding the package.
 - c. Give the Floor and Room Number (if needed) to be as descriptive as possible where this package needs to be picked up from. Click Save.

8. Print the airbill that is automatically downloaded.
 - a. To reprint airbill, click History at the top left of the page.
 - i. Shipments created from the user that day will automatically populate. If shipments from a previous day need to be located, search by ship date.
 - ii. Locate the correct shipment, and click on the printer icon to the left of the tracking number under “Action” to reprint the airbill
 - iii. Click print icon on right side of the tracking number line.

9. Fold airbill, and place inside plastic UPS sleeve. Peel the back off of the UPS sleeve and stick the sleeve to the top of the package. Do NOT overlap other labels or package seams.

7.3 Frozen Shipment Notification Electronic Data Capture Submission Instructions

2. Submit the electronic shipment notification form by clicking here: <https://redcap.link/NCRADShippingForm> This form must be completed **prior to each NCRAD shipment.**
3. Enter all required information, including:
 - a. Contact information
 - b. Shipping information
 - c. Kit numbers
 - d. Participant IDs
4. Upload **one (1) zip file** that contains all saved *PDF Condensed Forms* for the participant samples included in the shipment.
5. Enter "Shipment Date".
6. Click "Submit".

8.0 Data Queries and Reconciliation

Sample and Shipment Notification forms must be completed on the day that samples are collected (for ambient samples), or before sample shipment (for batch frozen samples), as these forms capture information that will be used to reconcile sample collection and receipt, as well as for future analyses.

All sample processing information must be submitted electronically using the NCRAD electronic Sample Form, available at: <https://redcap.uits.iu.edu/MCSSampleForm> promptly after processing samples. Upon submission, a copy of the completed Sample Form and a Condensed Form will be automatically emailed to the submitter. The Condensed Form must be saved as a *PDF* and a *physical copy printed*.

In addition, the electronic Sample Shipment Notification Form must be completed prior to each UPS shipment using the following link: <https://redcap.link/NCRADShippingForm> When completing the electronic shipment notification form, a single ZIP file must be uploaded that contains the *PDF copies* of all saved Condensed Forms for the participant samples included in the shipment. A *printed physical copy* of the Condensed Sample Form for each participant must be placed inside the UPS shipping box, positioned on top of the Styrofoam container, with each shipment.

NCRAD will collaborate with the data team at NACC to reconcile information captured in the NACC database compared to samples received and logged at NCRAD. Additional discrepancies may be sent directly to the Center staff to reconcile.

Data queries or discrepancies with samples shipped and received at NCRAD may result from:

- Incorrect samples collected and shipped
- Damaged or incorrectly prepared samples
- Unlabeled samples, samples labeled with incomplete information, or mislabeled samples
- Discrepant information documented on the Blood Sample and Shipment Notification Form and logged at NCRAD compared to information entered into the NACC database.

9.0 Appendices

[Appendix A: Rate of Centrifuge Worksheet](#)

[Appendix B: Blood Sample and Shipment Notification Form](#)



Biospecimen Collection, Processing, and Shipment Manual
Appendix A: Rate of Centrifuge Worksheet

Please complete and return this form by fax or email to the NCRAD Project Manager if you have any questions regarding sample processing. The correct RPM will be sent back to you.

Submitter Information

Name:

Site:

Submitter e-mail:

Centrifuge Information

Please answer the following questions about your centrifuge.

Centrifuge Type

Fixed Angle Rotor:

Swing Bucket Rotor:

Radius of Rotation (mm):

Determine the centrifuge’s radius of rotation (in mm) by measuring distance from the center of the centrifuge spindle to the bottom of the device when inserted into the rotor (if measuring a swing bucket rotor, measure to the middle of the bucket).

Calculating RPM from G-Force:

$$RCF = \left(\frac{RPM}{1,000} \right)^2 \times r \times 1.118 \Rightarrow RPM = \sqrt{\frac{RCF}{r \times 1.118}} \times 1,000$$

RCF = Relative Centrifugal Force (G-Force)

RPM = Rotational Speed (revolutions per minute)

R= Centrifugal radius in mm = distance from the center of the turning axis to the bottom of centrifuge

Comments:

Please send this form to NCRAD Study Coordinator

317-321-2003 (Fax)

alzstudy@iu.edu



Appendix B: Blood Sample and Shipment Notification Form

Please email or fax the form on or prior to the date of shipment.

To: Kelley Faber Email: alzstudy@iu.edu Phone: 1-800-526-2839/317-278-8413

From: Mahesh Joshi, PhD UPS tracking #: 1Z976R8W84

Phone: 561-869-6827 Email: maj145@med.miami.edu

Study: Multicultural Community Dementia Screening

Visit: Baseline 1-Year 2-Year 3-Year 4-Year 5-Year

KIT BARCODE

Sex: M F **Year of Birth:** _____

Participant ID: HBI _____

Blood Collection:

Date of Draw: _____ [MMDDYY]	Time of Draw: _____ [HHMM]
Date participant last ate: _____ [MMDDYY]	Time participant last ate: _____ [HHMM]

Blood Processing:

Plasma & Buffy Coat (EDTA Tube)

Original blood volume of EDTA #1: _____	Original blood volume of EDTA #2: _____ mL
Time spin started: _____ [HHMM]	Duration of centrifuge: _____ mins
Temp of centrifuge: _____ °C	Rate of centrifuge: _____ x g
Time aliquoted: _____ [HHMM]	Number of 1.5 mL plasma aliquots created (purple cap, up to 6): _____
If applicable, volume of residual plasma aliquot (less than 1.5 mL in blue cap): _____ mL <input type="checkbox"/>	If applicable, specimen number of residual plasma aliquot (Last four digits): _____ <input type="checkbox"/> N/A
Buffy coat #1 specimen number (Last four digits): _____	Buffy coat #1 volume: _____ mL
Buffy coat #2 specimen number (Last four digits): _____	Buffy coat #2 volume: _____ mL
Time aliquots placed in freezer: _____ [HHMM]	Storage temperature of freezer: _____ °C

Notes: _____